

Recovering Microalgae Using a Salsnes Filter

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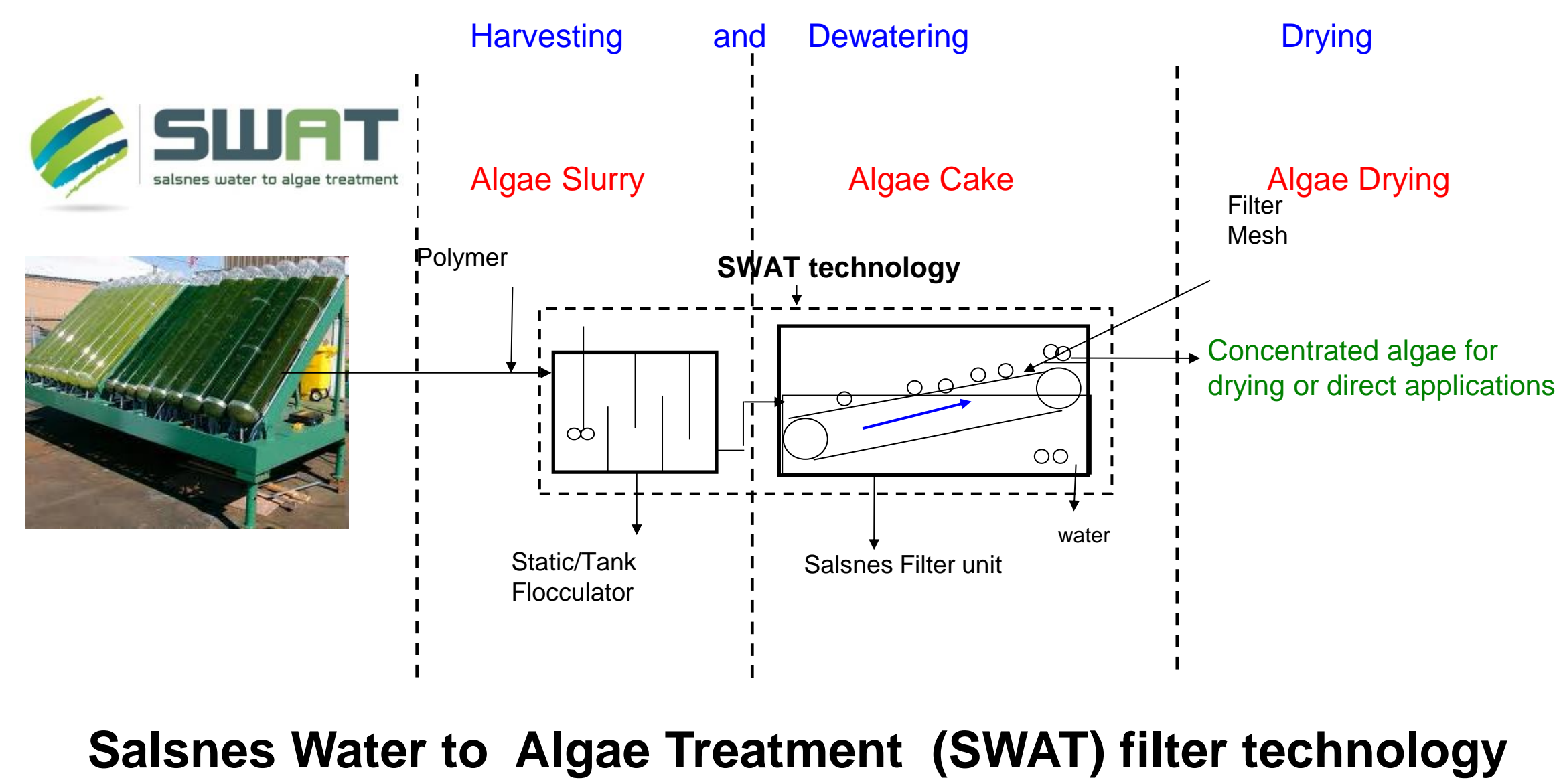
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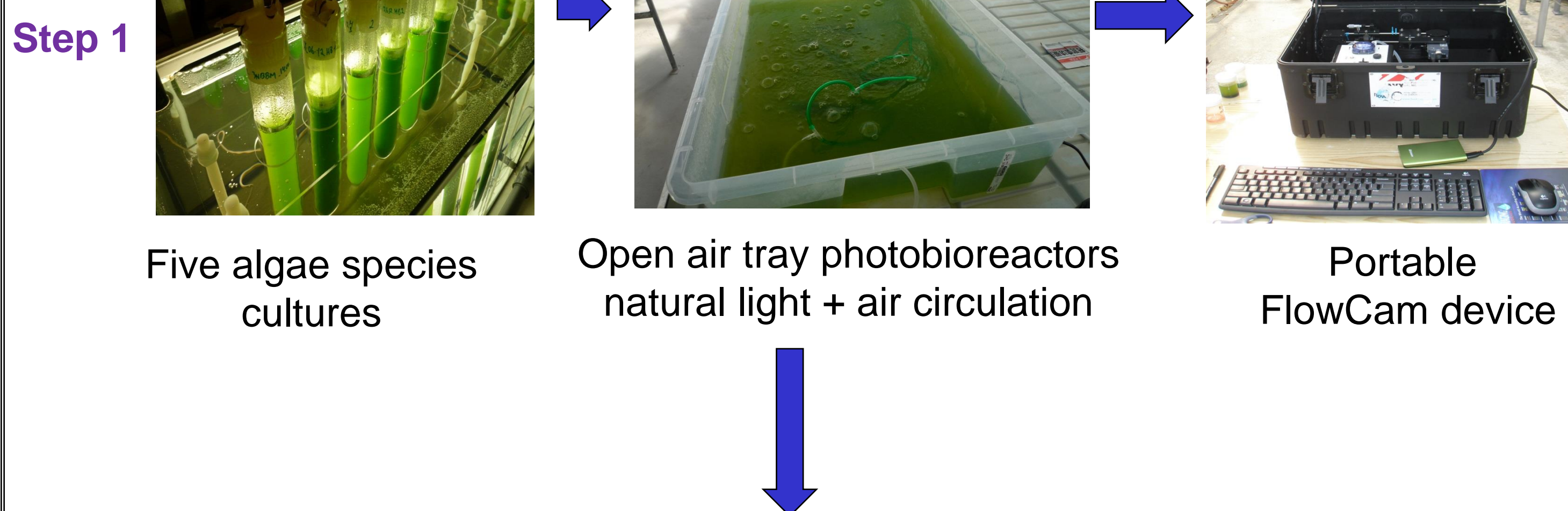
Introduction & Objectives

- The overall goal is to develop a universal microalgae harvesting technology:
 - by building on the experiences gained from removal of particles from wastewater
 - by modifying the current wastewater technologies such as Salsnes Filter (SF)
- SF has been successfully used for primary treatment of municipal wastewater
- Aim is to recover >95% of microalgae from water phase and use <0.08 kWh/m³ of algae



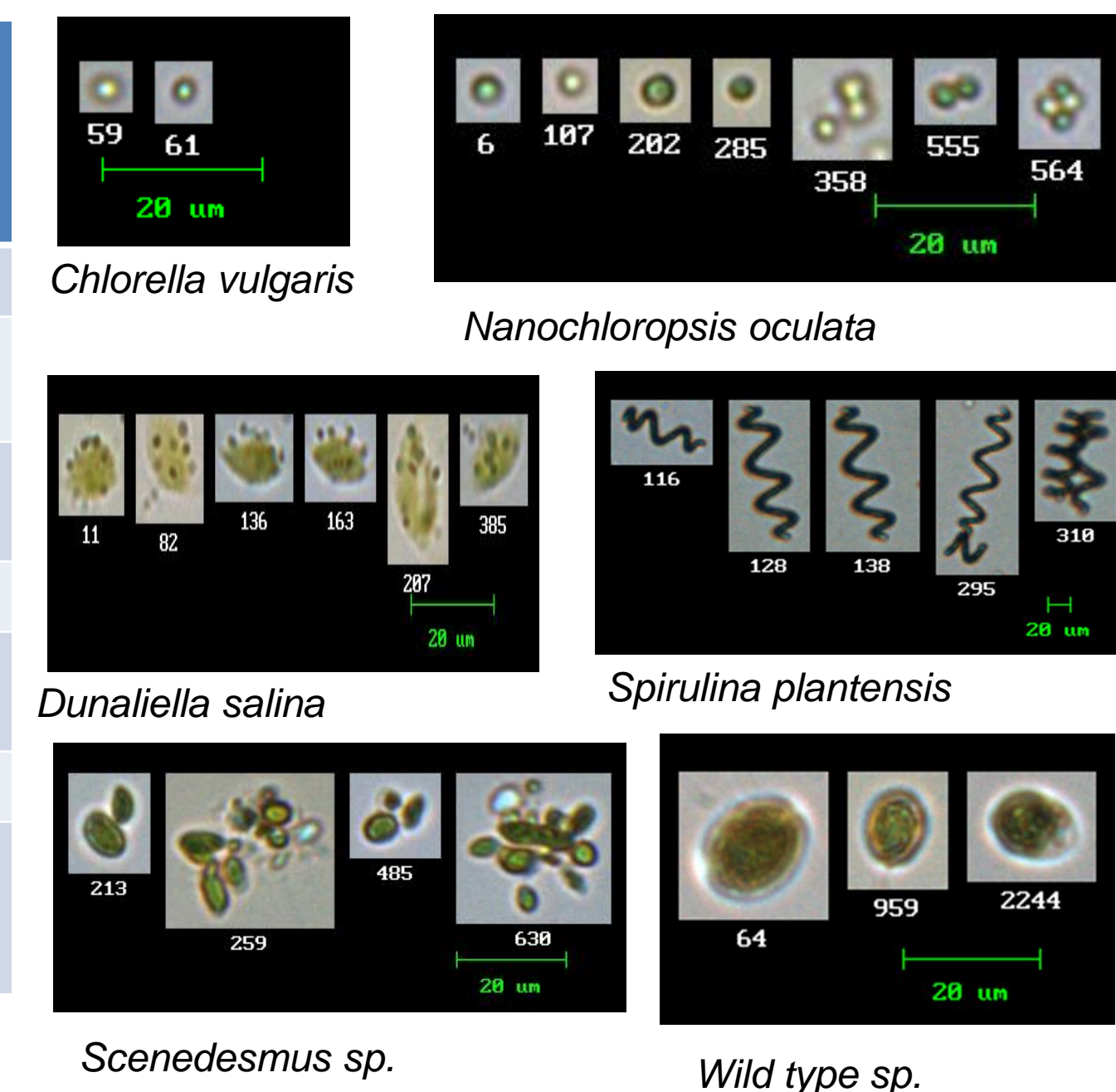
Methods, Materials, Results

Culture, grow 5 species, evaluate cell size, growth conditions

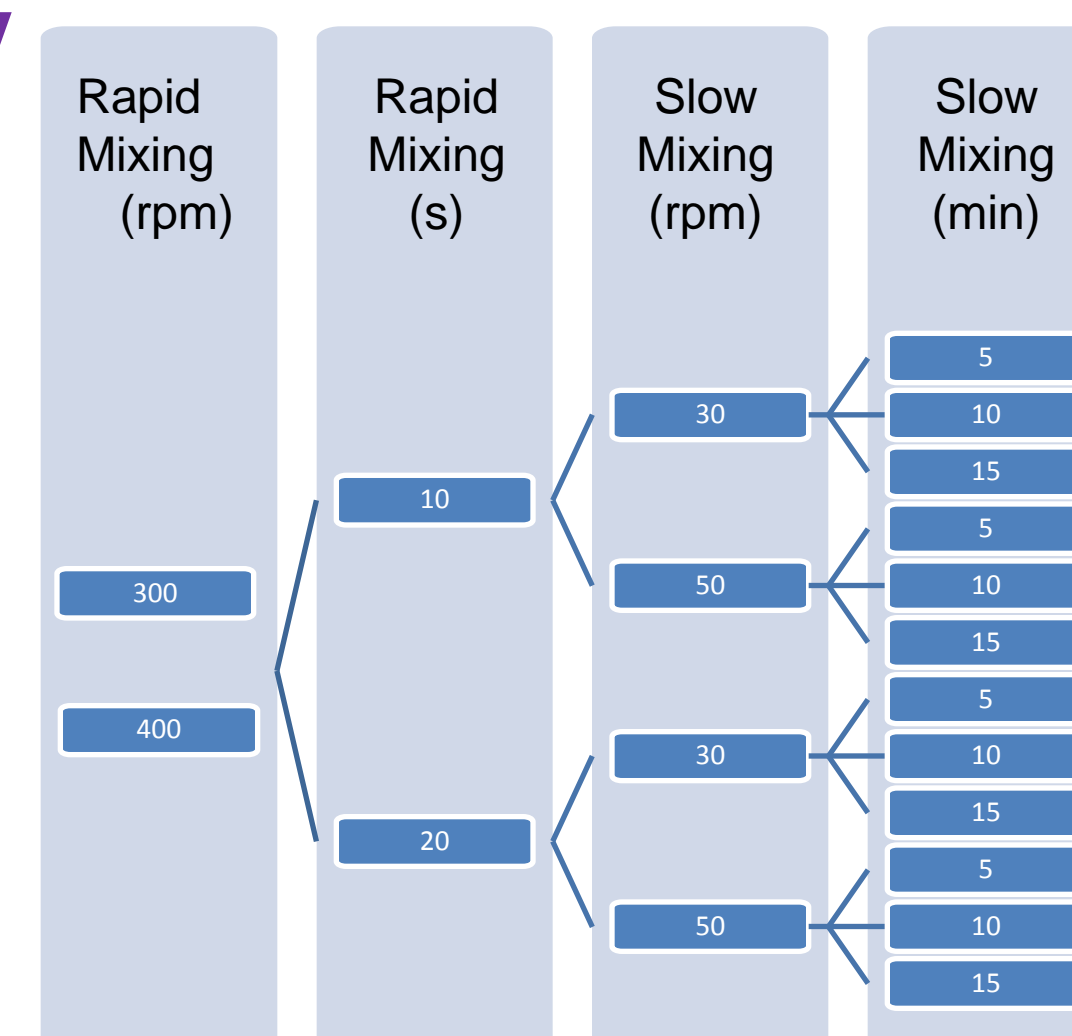
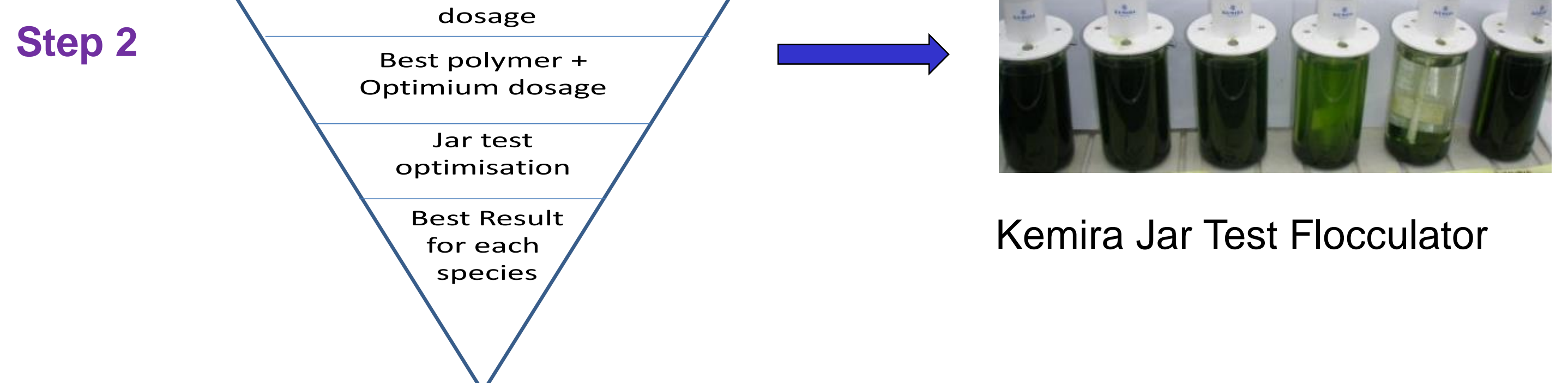


Species	Shape	Media	Avg particle size (L)	Number of particles analysed	Area based diameter
			µm	#	µm
<i>Chlorella vulgaris</i>	Spherical	FW	5.07	3838	3.99±1.26
<i>Dunaliella salina</i>	Irregular	M	7.26	646	4.84±3.7
<i>N. Oculata</i>	Spherical	M	4.8	3805	3.76±0.94
<i>Scenedesmus sp</i>	Rod	FW	20.01	440	14.7±6.61
<i>Wild type</i>	Oval	WW	50.43	54	14.8±7.83
<i>Spirulina plantensis</i>	Spiral	FW	111.99	3179	112± 80 (L) 47±36 (W)

FW- fresh water; M-Marine; WW-Wastewater



Test different polymers + chemicals, determine mixing intensity



Species	% Removal of Turbidity	Rapid mixing (300 rpm)	Slow mixing (30-50 rpm)
		s	min
<i>Chlorella vulgaris</i>	95	10	5
<i>Dunaliella salina</i>	98.8	10	5
<i>N. Oculata</i>	79	20	10
<i>Scenedesmus sp</i>	98.9	20	5
<i>Wild type</i>	64	10	10

Test different size flocculators, using scaled up G values



The power requirements were calculated using Equation 1. This equation was introduced by Camp and Stein (1943) as a measurable average value to replace the local velocity gradient during turbulent mixing.

$$G = \sqrt{\frac{P}{\mu V}} \quad (\text{Equation 1})$$

where

- P energy dissipation from mixing (W)
- μ absolute viscosity of the liquid
- V volume of the tank (m³)

For a mechanical mixer, the power consumed by the mixer is given by (Bratby, 2006)

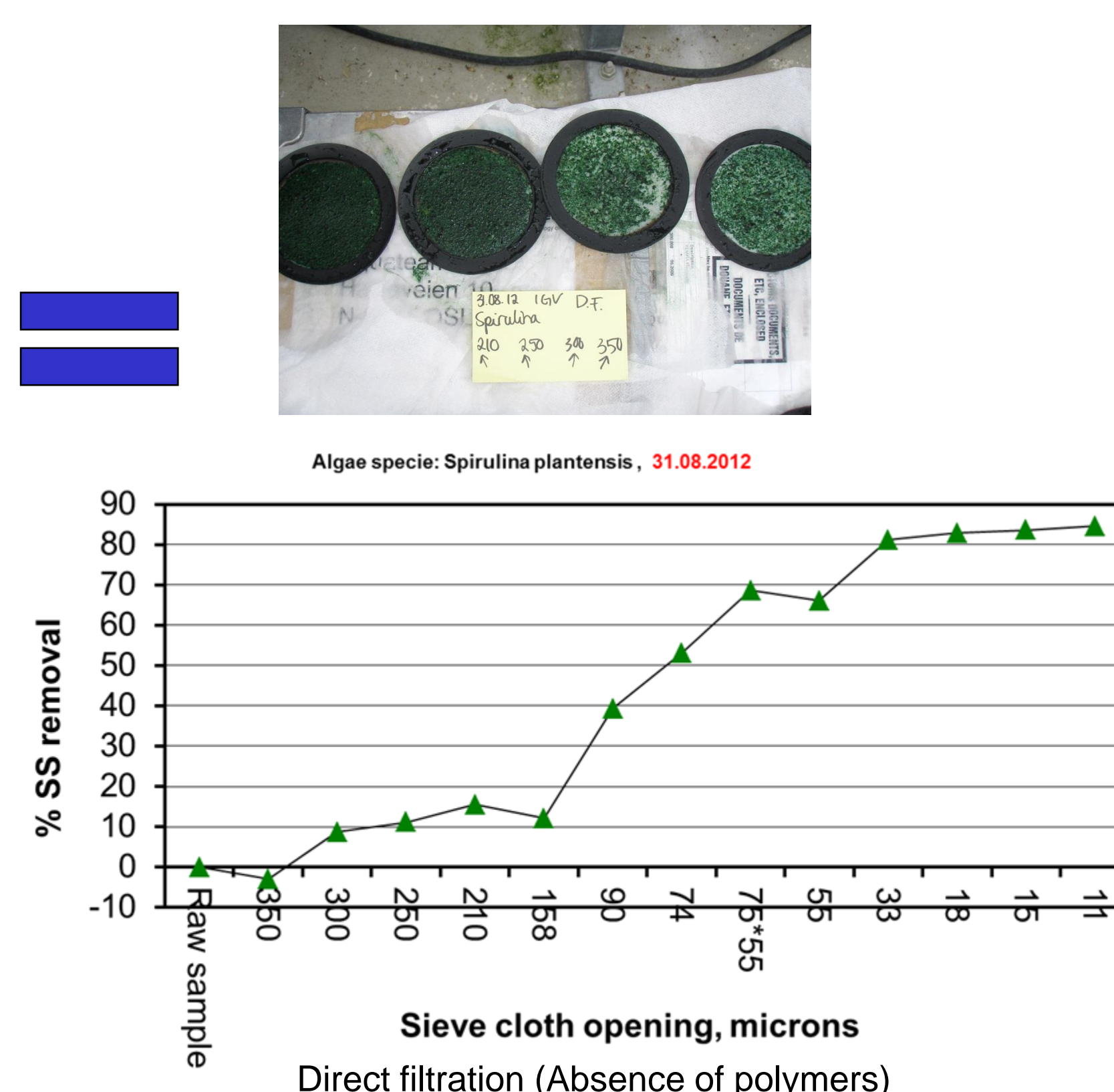
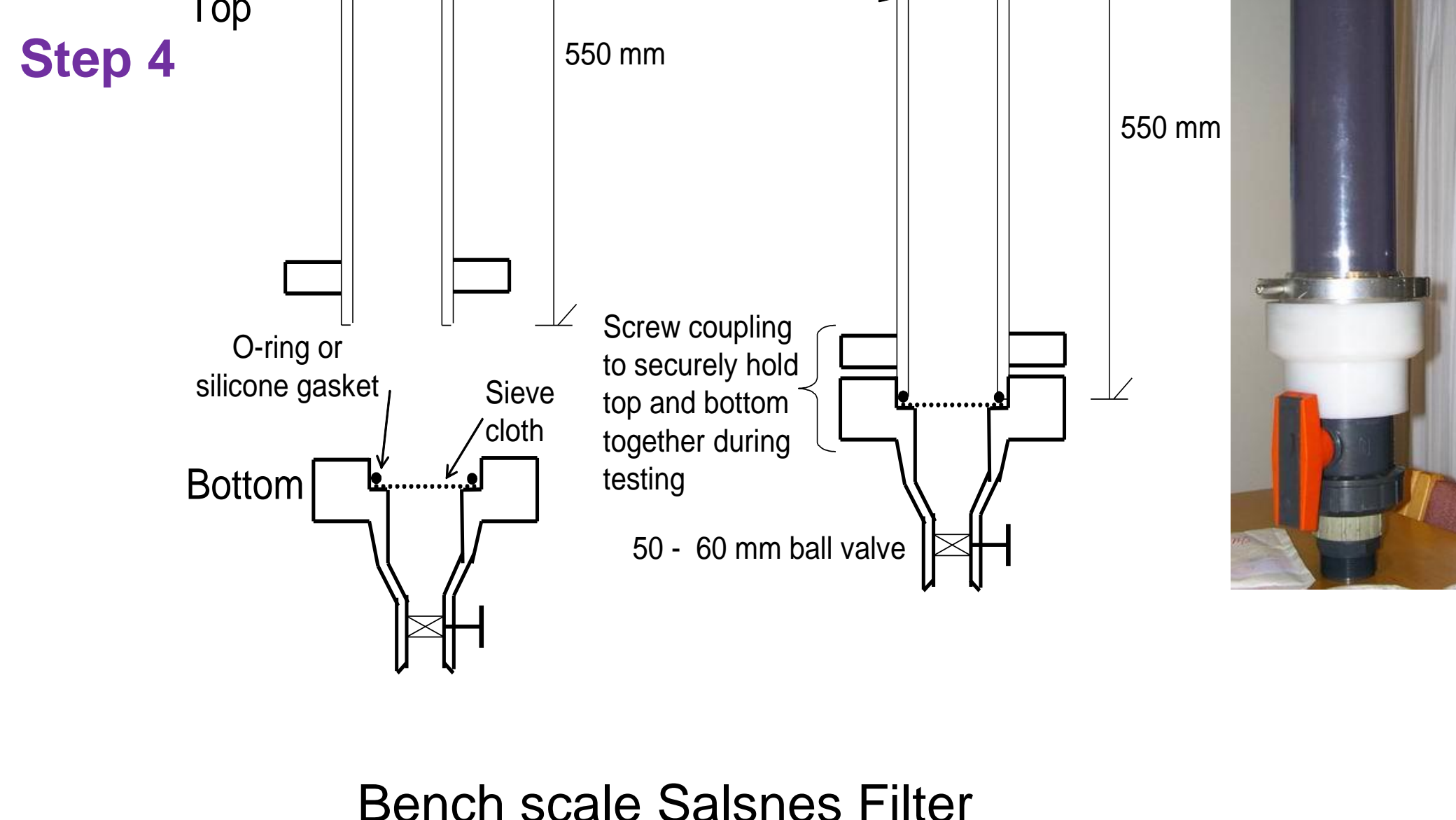
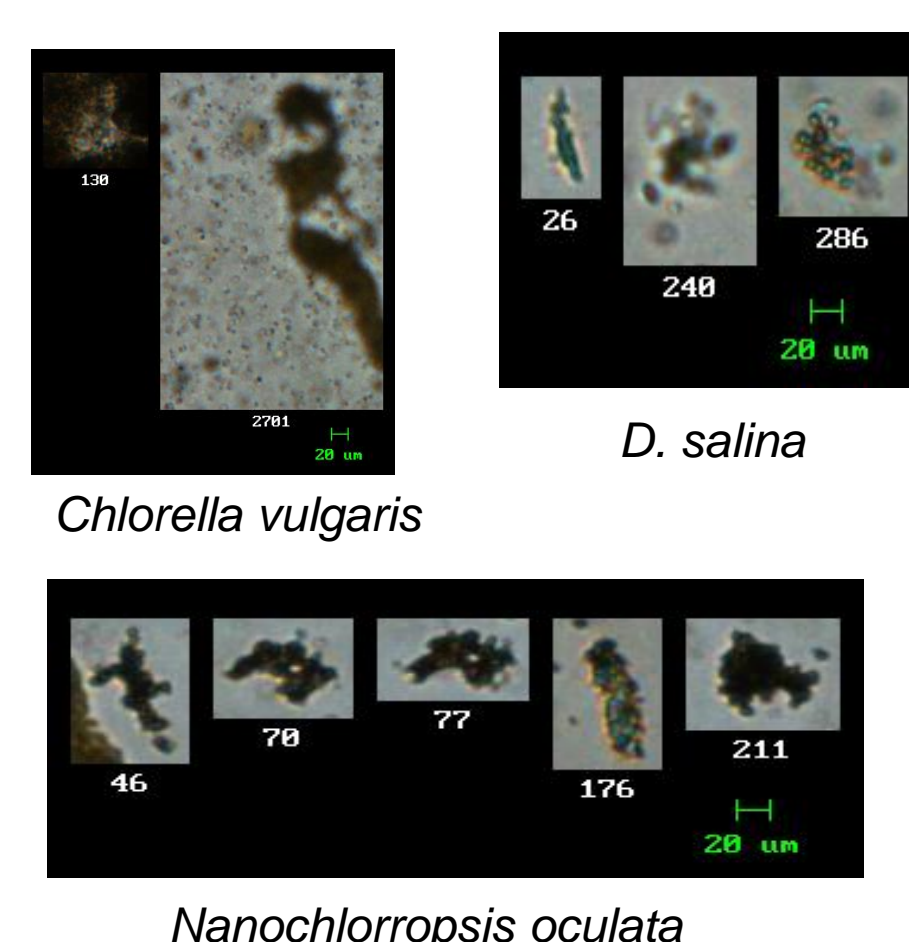
$$P = \phi \cdot \rho \cdot n^3 \cdot D^5 \quad (\text{Equation 2})$$

where

- φ dimensionless power number
- ρ liquid density (kg/m³)

	Rapid Mixing		Slow Mixing	
	rpm	s	rpm	Min
Jar Test	300	20	50	5
20 L	232	20	74	5
BSAF	95	20	30	5

Species	Avg floc size (L)	Number of flocs analysed	Area based diameter range
	µm	#	Sq. µm
<i>Chlorella vulgaris</i>	181.40	227	72.46±14.24
<i>Dunaliella salina</i>	71.35	428	41.91±9.85
<i>N. Oculata</i>	59.05	2650	35.29±8.66
<i>Scenedesmus sp</i>	78.21	4499	40.73±12.38
<i>Wild type</i>	90.96	892	52.28±14.49



- Direct filtration results clearly showed that the tested algae species need to be flocculated in to larger particles in order to achieve the goal of 90% recovery when harvested on a filter mesh sieve
- Optimum flocculant and optimum dose is very dependent on the type of algae
- Very good flocculation could be achieved with all the tested algae species, but for some species it may not be achieved at an economically acceptable chemical dose
- Bench scale flocculation and Salsnes filtration resulted in 93 % and 96 % recovery of *Chlorella vulgaris* and *Scenedesmus sp*, respectively. Results based on TSS
- Pilot scale flocculation (20 L) and Salsnes filtration resulted in 92 % and 84% recovery of *Nanochloropsis oculata* and *Wild type sp*, respectively. Results based on TSS.



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